

PRODUCT MONOGRAPH

PrNOVO-CEFUROXIME

(Cefuroxime Axetil Tablets, USP)

125 mg, 250 mg and 500 mg cefuroxime/tablet

Antibiotic

Control Number: 031000
Novopharm Limited
Toronto, Canada

Date of Preparation:
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ACTIONS AND CLINICAL PHARMACOLOGY

Cefuroxime axetil is an orally active prodrug of cefuroxime. After oral administration, NOVO-CEFUROXIME (cefuroxime axetil) is absorbed from the gastrointestinal tract and rapidly hydrolyzed by nonspecific esterases in the intestinal mucosa and blood to release cefuroxime into the blood stream. Conversion to cefuroxime, the microbiologically active form, occurs rapidly. The inherent properties of cefuroxime are unaltered after its administration as cefuroxime axetil. Cefuroxime exerts its bactericidal effect by binding to an enzyme or enzymes referred to as penicillin-binding proteins (PBPs) involved in bacterial cell wall synthesis.

This binding results in inhibition of bacterial cell wall synthesis and subsequent cell death. Specifically, cefuroxime shows high affinity for PBP 3, a primary target for cefuroxime in gram-negative organisms such as *E. coli*.

A comparative, two-way, single-dose, fasting bioavailability study was performed on two 250 mg cefuroxime axetil products, NOVO-CEFUROXIME 250 mg tablets and Ceftin[®] 250 mg tablets. The pharmacokinetic data calculated for cefuroxime axetil in the NOVO-CEFUROXIME and Ceftin[®] tablet formulations is tabulated in Table 1.

Table 1

| | Geometric Mean Arithmetic Mean (C.V.) | | Percentage of Ceftin [®] |
|--------------------------------|--|--|-----------------------------------|
| | Novo-Cefuroxime (2 x 250 mg) | Ceftin [®] ** (2 x 250 mg) | |
| AUC _T (ng•hr/mL) | 15.65 16.33 (28) | 16.44 17.48 (36) | 95 |
| AUC _I (ng•hr/mL) | 16.36 16.97 (27) | 17.12 18.16 (35) | 96 |
| C _{max} (ng/mL) | 5.39 5.63 (28) | 5.50 5.76 (32) | 98 |

| | | | |
|----------------------------|-------------|-------------|-----|
| T _{max} * (hr) | 1.33 (0.25) | 1.74 (0.97) | --- |
| T _{1/2} * (hr) | 1.26 (0.20) | 1.32 (0.19) | --- |

*For the T_{max} and T_{1/2} parameters these are the arithmetic means (standard deviation).

**Ceftin® manufactured by Glaxo Canada Inc., Mississauga, Canada.

INDICATIONS AND CLINICAL USE

NOVO-CEFUROXIME (cefuroxime axetil) is indicated for the treatment of patients with mild to moderately severe infections caused by susceptible strains of the designated organisms in the following diseases:

Upper Respiratory Tract Infections

Pharyngitis and tonsillitis caused by *Streptococcus pyogenes*. Otitis Media caused by *Streptococcus pneumoniae*, *Streptococcus pyogenes* (group A beta-hemolytic streptococci), *Haemophilus influenzae* (beta-lactamase negative and beta-lactamase positive strains) or *Moraxella catarrhalis*. Sinusitis caused by *Moraxella catarrhalis*, *Streptococcus pneumoniae* or *Haemophilus influenzae* (including ampicillin-resistant strains).

Lower Respiratory Tract Infections

Pneumonia or bronchitis caused by *Streptococcus pneumoniae*, *Haemophilus influenzae* (including ampicillin-resistant strains), *Haemophilus parainfluenzae*, *Klebsiella pneumoniae* or *Moraxella catarrhalis*.

Skin Structure Infections

Skin structure infections caused by *Staphylococcus aureus*, *Streptococcus pyogenes* or *Streptococcus agalactiae*.

Gonorrhea

Acute uncomplicated urethritis and cervicitis caused by *Neisseria gonorrhoeae*.

Bacteriologic studies to determine the causative organism and its susceptibility to cefuroxime should be performed. Once these results become available antibiotic treatment should be adjusted if required.

CONTRAINDICATIONS

NOVO-CEFUROXIME (cefuroxime axetil) is contraindicated for patients who have shown Type 1 hypersensitivity to cefuroxime, to any of its components, or to any of the cephalosporin group of antibiotics.

WARNINGS

Before therapy with NOVO-CEFUROXIME (cefuroxime axetil) is instituted, careful inquiry should be made to determine whether the patient has had previous hypersensitivity reactions to cefuroxime, cephalosporins, penicillin, or other drugs. NOVO-CEFUROXIME should be administered with caution to any patient who has demonstrated some form of allergy, particularly to drugs. There is some clinical and laboratory evidence of partial cross-allergenicity of the cephalosporins and penicillin. Special care is indicated in patients who have experienced anaphylactic reaction to penicillins or other beta-lactams. If an allergic reaction to NOVO-CEFUROXIME occurs, treatment should be discontinued and standard agents (e.g. epinephrine, antihistamines, corticosteroids) administered as necessary.

Pseudomembranous colitis has been reported to be associated with the use of cefuroxime axetil and other broad-spectrum antibiotics. Therefore, it is important to consider its diagnosis in patients administered NOVO-CEFUROXIME who develop diarrhea. Treatment with broad spectrum antibiotics, including cefuroxime axetil, alters the normal flora of the colon and may permit overgrowth of *Clostridia*. Studies indicate that a toxin produced by *Clostridium difficile* is one primary cause of antibiotic associated colitis. Mild cases of colitis may respond to drug discontinuance alone. Moderate to severe cases should be managed with fluid, electrolyte, and protein supplementation as indicated. When the colitis is severe or not relieved by discontinuance of NOVO-CEFUROXIME administration, consideration should be given to the administration of oral vancomycin or other suitable therapy. Other possible causes of colitis should also be considered.

PRECAUTIONS

General

Broad-spectrum antibiotics including NOVO-CEFUROXIME (cefuroxime axetil) should be administered with caution to individuals with a history of gastrointestinal disease, particularly colitis.

The concomitant administration of aminoglycosides and some cephalosporins has caused nephrotoxicity. There is no evidence that cefuroxime, when administered alone, is nephrotoxic, although transient elevations of BUN and serum creatinine have been observed in clinical studies. However, the effect of administering NOVO-CEFUROXIME concomitantly with aminoglycosides is not known.

Studies suggest that the concomitant use of potent diuretics, such as furosemide and ethacrynic acid, may increase the risk of renal toxicity with cephalosporins.

As with other antibiotics, use of NOVO-CEFUROXIME may result in the overgrowth of *Candida*. Prolonged use may also result in the overgrowth of other non-susceptible organisms (e.g. enterococci and *Clostridium difficile*), which may require interruption of treatment.

Repeated evaluation of the patient's condition is essential. If superinfection occurs during therapy, appropriate measures should be taken. Should an organism become resistant during antibiotic therapy, NOVO-CEFUROXIME should be discontinued and another appropriate antibiotic should be substituted.

Pregnancy

The safety of NOVO-CEFUROXIME in pregnancy has not been established. The use of cefuroxime axetil in pregnant women requires that the likely benefit from the drug be weighed against the possible risk to the mother and fetus. Animal studies following parenteral administration have shown cefuroxime to affect bone calcification in the fetus and to cause maternal toxicity in the rabbit. Reproduction studies that have been performed in mice and rats at oral doses of up to 50 to 160 times the human dose have revealed no evidence of impaired fertility or harm to the fetus due to cefuroxime axetil. There are, however, no adequate and well-controlled studies in pregnant women. Because animal reproduction studies are not always predictive of human response, this drug should be used during pregnancy only if clearly needed.

Nursing Mothers

Since cefuroxime is excreted in human milk, consideration should be given to discontinuing nursing temporarily during treatment with NOVO-CEFUROXIME.

Drug Interactions

Drugs which reduce gastric acidity may result in a lower bioavailability of NOVO-CEFUROXIME compared with that of the fasting state and tend to cancel the effect of enhanced post-prandial absorption. In common with other antibiotics, cefuroxime axetil may affect the gut flora, leading to lower estrogen reabsorption and reduced efficacy of combined oral contraceptives.

Drug-Laboratory Test Interactions

A false-positive reaction for glucose in the urine may occur with copper reduction tests (Benedict's or Fehling's solution or with Clinitest[®] Tablets) but not with enzyme-based tests for glycosuria (e.g. Clinistix[®], Tes-Tape[®]). As a false-negative result may occur in the ferricyanide test, it is recommended that either the glucose oxidase or hexokinase method be used to determine blood plasma glucose levels in patients receiving NOVO-CEFUROXIME .

Cefuroxime does not interfere with the assay of serum and urine creatinine by the alkaline picrate method.

Cephalosporins as a class tend to be absorbed onto the surface of red cell membranes and react with antibodies directed against the drug to produce a positive Coombs' test (which can interfere with cross-matching of blood) and very rarely hemolytic anemia.

Ability to Perform Tasks That Require Judgement, Motor or Cognitive Skills

As this medicine may cause dizziness, patients should be warned to be cautious when driving or operating machinery.

ADVERSE REACTIONS

The following adverse reactions have been reported:

Gastrointestinal (approximately 8% of the patients): Diarrhea (5.6%), nausea (2.4%), vomiting (2.0%), loose stools (1.3%). Reports abdominal pain have occurred.

Hepatic (3% of patients): Transient increases of hepatic enzyme levels [ALT (SGPT), AST (SGOT), LDH].

Central Nervous System (2.2% of patients): Headache and dizziness.

Hypersensitivity (1.3% of patients): Rashes (0.6%), pruritus (0.3%), urticaria (0.2%), shortness of breath and rare reports of bronchospasm. Hypersensitivity reactions to NOVO-CEFUROXIME (cefuroxime axetil) may occur in patients who report delayed hypersensitivity to penicillins (see WARNINGS). As with other cephalosporins, there have been rare reports of drug fever.

Hematologic: increased erythrocyte sedimentation rate, eosinophilia, decreased hemoglobin and very rarely hemolytic anemia.

Miscellaneous: The following adverse reactions have been observed to occur, although infrequently, in association with parenteral cefuroxime sodium and may be potential adverse effects of oral cefuroxime axetil: drowsiness, vaginitis, positive direct Coombs test, and transient increases in serum bilirubin, creatinine, alkaline phosphatase, and urea nitrogen (BUN).

POSTMARKETING EXPERIENCE WITH CEFUROXIME PRODUCTS

In addition to adverse events reported during clinical trials, the following events have been identified during clinical practice in patients treated with cefuroxime axetil tablets and were reported spontaneously. Data are generally insufficient to allow an estimate of incidence or to establish causation.

General: The following hypersensitivity reactions have been reported: anaphylaxis, angioedema, pruritus, rash, serum sickness-like reaction, urticaria.

Gastrointestinal: Pseudomembranous colitis (see WARNINGS).

Hematologic: thrombocytopenia, and leucopenia (sometimes profound).

Hepatic: Jaundice (predominantly cholestatic) and hepatitis have been reported very rarely.

Infections and Infestations: Candida overgrowth.

Neurologic: Seizure.

Skin: Erythema multiforme, Stevens-Johnson syndrome, toxic epidermal necrolysis.

Urologic: Renal dysfunction.

SYMPTOMS AND TREATMENT OF OVERDOSAGE

Other than general supportive treatment, no specific antidote is known. Excessive serum levels of cefuroxime can be reduced by dialysis. For treatment of hypersensitive reactions, See WARNINGS.

For management of a suspected drug overdose, contact your regional Poison Control Centre.

DOSAGE AND ADMINISTRATION

NOVO-CEFUROXIME (cefuroxime axetil) may be given orally without regard to meals. Absorption is enhanced when cefuroxime axetil is administered with food.

Adults and Children 12 Years of Age and Older:

The usual recommended dosage is 250 mg twice a day. However, dosage may be modified according to the type of infection present as indicated below:

| TYPE OF INFECTION | DOSAGE |
|--|---------------------|
| Pharyngitis, tonsillitis, sinusitis, bronchitis, skin structure infections | 250 mg twice daily |
| more severe infections eg. pneumonia | 500 mg twice daily |
| uncomplicated gonorrhea | 1000 mg single dose |

There is presently no data available on the effects of cefuroxime axetil in patients with renal impairment. However, in patients where there is significant impairment, a reduction in NOVO-CEFUROXIME dosage may be required.

Infants and Children less than 12 Years of Age:

NOVO-CEFUROXIME tablets are not recommended for infants and children less than 12 years of age.

The usual duration of treatment for NOVO-CEFUROXIME tablets is 7 to 10 days. For β -hemolytic streptococcal infections, therapy should be continued for at least 10 days.

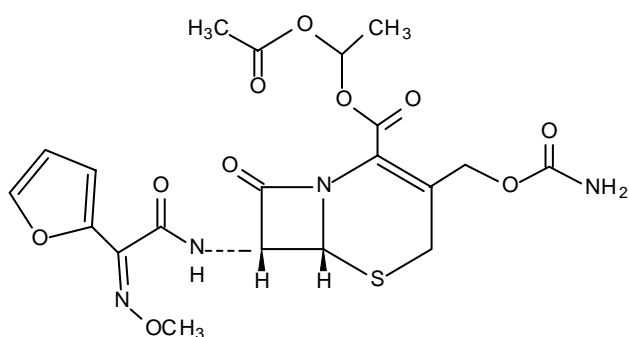
PHARMACEUTICAL INFORMATION

DRUG SUBSTANCE:

Proper Name: Cefuroxime Axetil

Chemical Name: 5-Thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid, 3-[[[(aminocarbonyl)oxy]methyl]-7-[[2-furanyl(methoxyimino)acetyl]amino]-8-oxo-, 1-(acetyloxy)ethyl ester, [6R-[6a,7b(Z)]]-

Structural Formula:



Molecular Formula: $C_{20}H_{22}N_4O_{10}S$

Molecular Weight: 510.7

Description: Cefuroxime axetil is an amorphous white to cream-coloured powder. It is soluble in dimethyl sulfoxide, dimethylformamide, 1,4-dioxan, chloroform, acetone, glacial acetic acid, ethyl acetate, and methanol. It is soluble with decomposition in 2N sodium hydroxide and slightly soluble in water, diethyl ether, 95% ethanol, and toluene, and insoluble in 2N hydrochloric acid. Cefuroxime axetil decomposes below its melting point.

STABILITY AND STORAGE

Store tablets between 15°C – 30°C

DOSAGE FORMS, COMPOSITION AND PACKAGING

NOVO– CEFUROXIME 125 mg tablets is available as a white to off-white, film coated, capsule shaped tablet engraved with novo on one side and 125 on the other. Each tablet contains cefuroxime axetil equivalent to 125 mg of cefuroxime.

NOVO– CEFUROXIME 250 mg tablets is available as a white, film coated, capsule shaped tablet engraved with novo on one side and 250 on the other. Each tablet contains cefuroxime axetil equivalent to 250 mg of cefuroxime.

NOVO– CEFUROXIME 500 mg tablets is available as a white, film coated, capsule shaped tablet engraved with novo on one side and 500 on the other. Each tablet contains cefuroxime axetil equivalent to 500 mg of cefuroxime.

NOVO– CEFUROXIME Tablets contain the following nonmedicinal ingredients: colloidal silicon dioxide, croscarmellose sodium, hydrogenated castor oil, hydroxypropyl methylcellulose, microcrystalline cellulose, propylene glycol and sodium lauryl sulfate.

NOVO-CEFUROXIME Tablets are supplied in bottles of 100, 500 and 1000.

INFORMATION FOR THE CONSUMER

What you should know about NOVO-CEFUROXIME Tablets

Please read this leaflet carefully before you start to take your medicine.

This provides a summary of the information available on your medicine. For further information or advice, ask your doctor or pharmacist.

1. The Name of Your Medicine

The name of your medicine is NOVO-CEFUROXIME Tablets. It contains cefuroxime axetil. This medicine is similar to other medicines called cephalosporins, which are antibiotics.

2. How to Obtain Your Medicine

NOVO-CEFUROXIME Tablets can only be obtained with a prescription from your doctor.

3. The Purpose of Your Medicine

Your doctor has prescribed NOVO-CEFUROXIME Tablets because you have an infection. NOVO-CEFUROXIME Tablets is used to kill the bacteria or "germs" which cause infections. The infection can be cleared up if you take your medication in the proper way.

4. Important Points to Note Before Taking Your Medicine

You should not use NOVO-CEFUROXIME Tablets if you are allergic to cephalosporins. Tell your doctor also if you are allergic to or react badly to penicillins or other antibiotics.

If testing urine for sugar, false positive reactions may occur if using methods dependent on copper reduction such as Fehling's or Benedict's solution or with Clinitest® Tablets. For this reason enzyme-based tests such as Tes-Tape® or Clinistix® should be used.

As this medicine may cause dizziness, you should be cautious when driving or operating machinery.

5. The Use of This Medicine During Pregnancy and Breast Feeding

Tell your doctor if you are pregnant or breast feeding a baby. If you are pregnant or breast feeding, your doctor may decide not to prescribe this medicine, although, there may be circumstances when your doctor advises you differently.

6. How to Take Your Medicine

You must take the medicine as prescribed by your doctor. If you are not sure how many tablets to take, or how often to take them, consult your doctor or pharmacist. **YOU SHOULD NOT INCREASE OR DECREASE THE PRESCRIBED DOSE UNLESS ADVISED BY YOUR DOCTOR.**

The usual dose for adults is one 250 mg tablet twice a day. NOVO-CEFUROXIME Tablets has a bitter taste, therefore, **do not chew or crush the tablets** but swallow each one whole with a drink of water.

NOVO-CEFUROXIME Tablets are more effective if taken after food.

The usual length of treatment is 7 - 10 days, although your doctor may adjust the prescription to suit your treatment. During the course of treatment, all the tablets must be taken to make sure that all germs have been killed. **CONTINUE TAKING THE TABLETS UNTIL THEY ARE FINISHED, EVEN IF YOU BEGIN TO FEEL BETTER.**

7. After Taking Your Medicine

If you experience wheeziness and tightness of chest, swelling of eyelids, face or lips, or develop skin lumps or hives, or a skin rash (red spots), tell your doctor immediately. Do not take any more medicine unless your doctor tells you to do so. He may decide to stop your treatment.

You may experience diarrhea, vomiting or symptoms that you do not understand. There is no need to stop taking your tablets, but you should tell your doctor of any of these symptoms as soon as possible.

If you feel worse or you have taken all the tablets and do not feel better **TELL YOUR DOCTOR AS SOON AS POSSIBLE.**

8. What to Do if an Overdose is Taken

It is important to follow the dosage instructions on the label of your medicine. Taking more than this dose is unlikely to be dangerous unless a large quantity of tablets or suspension is taken all at once. In this case, contact your doctor or nearest hospital emergency department immediately.

9. Storing Your Medicine

Keep your tablets in a safe place where children cannot reach them.

10. What to Do if You Miss a Dose

If you forget to take a dose, take another as soon as possible. Then continue with the normal dose. Do not double doses.

11. What to Do When You Stop Your Medicine

If your doctor decides to stop the treatment, do not keep any left over medicine unless your doctor tells you to. Please discard all unused NOVO-CEFUROXIME Tablets.

12. What is in Your Medicine

NOVO-CEFUROXIME Tablets are supplied in three strengths, containing 125 milligrams of cefuroxime (as cefuroxime axetil), 250 milligrams of cefuroxime (as cefuroxime axetil) or 500 milligrams of cefuroxime (as cefuroxime axetil). Your doctor will decide which strength you need.

REMEMBER: This medicine is for you. Only a doctor can prescribe it for you. Never give it to someone else. It may harm them even if their symptoms are the same as yours.

13. Further Information

This leaflet does not contain the complete information about your medicine. If you have any questions you should ask your doctor or pharmacist. You may need to read this leaflet again. Please do not throw it away until you have finished your medicine.

MICROBIOLOGY

The minimum inhibitory concentrations (MIC₅₀ and MIC₉₀) against various susceptible and non-susceptible organisms *in vitro* are shown in Table 2.

Table 2: Summary of *in vitro* activities against various organisms

| Organism (No. of Strains) | Range | MIC (µg/mL) ^a | | Reference |
|--|--------------|--------------------------|------|------------------------|
| | | 50% | 90% | |
| Haemophilus influenzae (97)* | 0.25-4 | 0.5 | 1 | Fass et al |
| Haemophilus parainfluenzae (2) | - | ≤ 0.06 | 0.25 | Knapp et al |
| Branhamella catarrhalis (53) | 0.125-1.0 | 0.5 | 1 | Alvarez et al |
| Neisseria meningitidis (40) | 0.06-0.25 | 0.12 | 0.12 | Trallero et al |
| Escherichia coli (229) | 0.5->64 | 1 | 4 | Knapp et al |
| Klebsiella pneumoniae (20) | 0.25->32 | 2 | 8 | Neu et al |
| Klebsiella oxytoca (20) | 2->64 | 2 | 16 | Neu et al |
| Citrobacter diversus (15) | 1->32 | 4 | 8 | Neu et al |
| Salmonella sp (21) | 0.5-4 | 1 | 4 | Neu et al |
| Shigella sp (24) | 0.25-32 | 1 | 4 | Neu et al |
| Proteus mirabilis (53) | 0.25->64 | 0.5 | 1 | Knapp et al |
| Staphylococcus aureus methicillin-susceptible (25) | 1-4 | N/A | 4 | Stratton et al |
| methicillin-resistant (25) | 2-64 | N/A | 64 | Stratton et al |
| Staphylococcus epidermidis methicillin-susceptible (25) | 0.5-2 | N/A | 2 | Stratton et al |
| methicillin-resistant (25) | 1-8 | N/A | 8 | Stratton et al |
| Staphylococcus saprophyticus (20) | 2-4 | 4 | 4 | Fass et al |
| Streptococcus pyogenes (18) | ≤ 0.015-0.06 | ≤ 0.015 | 0.06 | Neu et al |
| Streptococcus pneumoniae (15) | 0.015-0.12 | 0.015 | 0.12 | Neu et al |
| Streptococcus viridans (27) | 0.12-8 | 0.5 | 4 | Neu et al |
| Streptococcus faecalis (21) | >32 | >32 | >32 | Neu et al |
| Peptostreptococcus sp (10) | 0.06-4 | 0.25 | 4 | Goldstein et al |
| Fusobacterium sp (18) | ≤ 0.03-0.5 | 0.25 | 0.5 | Goldstein et al |
| Bacteroides sp (17) | 0.06-32 | 0.25 | 32 | Goldstein et al |
| Campylobacter pylori (30) | 0.01-0.25 | 0.125 | 0.25 | Garcia-Rodriguez et al |

*50 β-Lactamase positive, 47 β-Lactamase negative

N/A - Not available

Stability to Beta-Lactamases:

Although cefuroxime is resistant to hydrolysis by most beta-lactamases, these enzymes from certain species (*Bacteroides fragilis*, *Enterobacter* and indole-positive *Proteus sp*) have been shown to cause hydrolysis. Table 3 shows the degree of resistance of cefuroxime to beta-lactamase inactivation.

Table 3: Hydrolysis of cefuroxime by a range of beta-lactamases

| Source of enzyme | Enzyme class | µg of cefuroxime hydrolyzed/minute |
|---------------------------------------|--------------|------------------------------------|
| Escherichia coli (R ⁺ tem) | III | <1 |
| E. coli (R ⁺ GN238) | V | 4.5 |
| E. coli D31 | I | <1 |
| Proteus mirabilis | III | <1 |
| Klebsiella aerogenes K1 | IV | 54 |
| Enterobacter cloacae P99 | I | <1 |
| Proteus vulgaris | I | <1 |
| Bacteroides fragilis 1600 | I | 112 |
| Psuedomonas aeruginosa 1822 | I | <1 |
| Bacillus cerus 659/H9 | | 72 |
| Staphylococcus aureus PC1* | | <1 |

* Activity is expressed as micrograms hydrolyzed per hour.

Susceptibility Testing:

The results of susceptibility testing, by either disk-diffusion or tube-dilution techniques, should be interpreted according to the criteria in Table 4.

Table 4: Susceptibility Testing

| | zone diameter (30 µg cefuroxime disk) | approximate MIC correlate |
|--|--|---------------------------|
| SUSCEPTIBLE (susceptible to the usual doses) | ≥23 mm | ≤ 4 µg/mL |
| MODERATELY SUSCEPTIBLE* (intermediate) | 15-22 mm | 8-16 µg/mL |
| RESISTANT | ≤ 14 mm | ≥ 32 µg/mL |
| <u>CONTROL STRAINS</u> | | |
| <i>S. aureus</i> ATCC 25923 | 27-35 mm | 0.5-2 µg/mL |
| <i>E. coli</i> ATCC 25922 | 20-26 mm | 2-8 µg/mL |

* Organisms that produce zones of 15 to 22 mm may be susceptible if the infection is confined to tissues and fluids (e.g. urine) in which high antibiotic concentrations are attained.

Only cefuroxime disks should be used, since cefuroxime has been shown by *in vitro* tests to be active against certain strains found resistant when other beta-lactamase disks are used.

PHARMACOLOGY

Human

The bioavailability of cefuroxime tablets was investigated in a six-way crossover study with each of 12 male adult volunteers receiving a single intravenous dose of cefuroxime sodium and five oral doses of cefuroxime axetil. Blood samples were collected at specified intervals for 12 hours and urine for 24 hours following each dose. The results of this study are presented in Table 5.

Table 5: Pharmacokinetics of cefuroxime axetil administered as cefuroxime tablets to adults

| DOSE (mg) | ROUTE | FASTED/ FED | PEAK SERUM CON- CENTRATION ($\mu\text{g/mL}$) | TIME TO PEAK (h) | AREA UNDER SERUM LEVEL- TIME CURVE mg.h/L | URINE RECOVERY 0-12 h (mg) | % DOSE RECOVERED IN URINE | % DOSE ABSORBED RELATIVE TO I.V. | HALF- LIFE (h) |
|-----------|-------|----------------|---|---------------------------|---|-------------------------------------|---------------------------------|---|-------------------|
| 500 | i.v. | fasted | 53.2 | 0.1 | 52.8 | 415 | 83 | 100 | 1.3 |
| 500 | po | fasted | 4.9 | 2.3 | 18.9 | 161 | 32 | 36 | 1.6 |
| 125 | po | fed | 2.1 | 2.2 | 6.7 | 65 | 52 | 51 | 1.2 |
| 250 | po | fed | 4.1 | 2.5 | 12.9 | 127 | 51 | 49 | 1.2 |
| 500 | po | fed | 7.0 | 3.0 | 27.4 | 242 | 48 | 52 | 1.2 |
| 1000 | po | fed | 13.6 | 2.5 | 50.0 | 434 | 43 | 47 | 1.3 |

The mean values of pharmacokinetic parameters after 12 volunteers received a single i.v. dose of cefuroxime and 5 oral doses of cefuroxime axetil.

Increasing doses of cefuroxime produced linear increases in peak serum concentrations and AUC.

Bioavailability appears to be independent of dose but is increased by the presence of food. Absolute bioavailability of cefuroxime tablets (500 mg dose) increased from 36% in fasted subjects to 52% after food.

The amount of cefuroxime excreted in the urine over 24 hours averaged 83% following intravenous dosing and ranged from 43% to 52% following oral dosing when taken after food.

The half-life of cefuroxime following oral administration as cefuroxime to healthy adult volunteers is 1.2 to 1.6 hours.

Animal Pharmacology

The secondary pharmacological effects of cefuroxime axetil have been investigated in mice, rats, and dogs following a single oral dose. Negative controls were administered a placebo suspension while positive controls received mecamlamine HCl (an inhibitor of gastrointestinal propulsion). The results are summarized in Table 6.

Table 6: Secondary pharmacological actions in animals after a single oral dose of cefuroxime axetil

| Animal | Dose (mg/kg) | No. of Animals* | Pharmacological Actions | Observation Times | Effects |
|--------|--------------|-----------------|---|-------------------------------|---------------------------------------|
| mice | 0.5 | 10 | pupil diameter, body temperature, gross behaviour | 0-1h, 24h intervals for 7 d | decreased body temperature in females |
| rat | 0.5 | 10 | pupil diameter, body temperature, gross behaviour | 0-1 h, 24 h intervals for 7 d | decreased body temperature in females |
| dog | 0.5 | 2 | BP, HR, ECG, gross behaviour | 2.25, 3, 6, 24 h | none |
| rat | 0.5 | 10 | gastrointestinal propulsion | 0.75h | none |

* Each group consisted of equal numbers of males and females

As can be seen, cefuroxime axetil has no effects on behaviour or pupil diameter in the mouse or the rat or on gastrointestinal propulsion in the rat. Cefuroxime axetil administered orally to dogs produced no acute effects on blood pressure, heart rate, or the electrocardiogram.

TOXICOLOGY

Acute Toxicity:

The experimental details of single-dose toxicity studies are presented in Table 7.

Table 7: Acute Toxicity

| Animal | Age | Route | Doses (g/kg) | Animals/Dose* | Length of Observation | LD ₅₀ (g/kg) |
|--------|-------|-------|--------------|---------------|-----------------------|-------------------------|
| mouse | adult | p.o. | 0, 6 | 20 | 3 days | > 6 |
| | adult | p.o. | 0, 6 | 20 | 14 days | > 6 |
| | adult | p.o. | 6 | 20 | 14 days | > 6 |
| | adult | p.o. | 1.5, 3 | 12 | 14 days | > 6 |

| | | | | | | |
|-----|---------|----------|--------|--------|---------|---------|
| rat | adult | p.o. | 0, 6 | 12 | 3 days | > 6 |
| | adult | p.o. | 0, 6 | 12 | 14 days | > 6 |
| | adult | -0 | 6 | 12 | 14 days | > 6 |
| | 10 days | p.o. | 0, 3 | 20 | 3 days | > 3 |
| | 10 days | p.o. | 0,3 | 20 | 14 days | > 3 |
| | adult | s.c. | 1.5, 3 | 12 | 14 days | > 3 |
| | dog | 8-10 mo. | p.o. | 1.5, 3 | 4 | 14 days |

* Each dosage group was composed of equal number of males and females

All animals survived the experiments. The median lethal dose of cefuroxime axetil in these animals is therefore, in excess of the test doses administered.

The only symptom of systemic toxicity observed was a temporary reduction in body weight in juvenile (10-day-old) rats. All other animals were found to be healthy during the observation period.

Mice and rats were sacrificed following the observation period. Histological examination of major organs and tissues revealed no evidence of systemic toxicity.

Subcutaneous injections in mice and rats caused local swelling due to the accumulation of large volumes of suspension. Small localized subcutaneous accumulations of test compound with slight peripheral tissue reaction were observed at autopsy.

Compacted test material was still present in the stomachs of most mice and rats three days after oral dosing. Inflammatory changes observed in the stomach walls of several animals were considered to be due to the mechanical irritation caused by these masses.

Long- Term Toxicity:

In subacute and chronic studies in rats receiving high oral doses of cefuroxime axetil (1.0-2.5 g/kg), accumulation of drug substance and the formation of concretions in the stomachs of many animals caused high rates of mortality. The concretions consisted of semicrystalline axetil, water, food, polymers and impurities comparable to that in the administered material.

Concretions were not a problem in the dog studies, which all proceeded to completion.

Experimental details of subacute and chronic toxicity studies are presented in Table 8.

Table 8: Subacute and Chronic Toxicity

| Animal | Ages* | Route | Daily Doses (g/kg) | Animals / Dose** | Intended Duration of Treatment Recovery | |
|--------|----------|-------|-------------------------|------------------|---|---------|
| rat | 7-9 wk | p.o. | 0, 0.1, 0.4, | 12 | 15 wk | - |
| | 7-9 wk | p.o. | | 12 | 15 wk | 22 days |
| | 7-9 wk | p.o. | 0.8, 1.7, 2.5 | 12 | 15 wk | - |
| rat | 8-10 wk | p.o. | 0, 0.1, 0.4, 1.6 | 60 | 28 wk | - |
| | 9 wk | p.o. | 0, 0.15, 0.4, 1.0 | 30 | 90 days | - |
| rat | 7 wk | p.o. | 0, 0.1, 0.4, 1.0 | 32 | 28 wk | - |
| | 7 wk | p.o. | | 24 | 28 wk | 5 wk |
| | 7 wk | p.o. | | 12 | 31 wk | - |
| dog | 12-16 wk | p.o. | 0, 0.1, 0.2 0.4, 0.8 | 6 | 5 wk | - |
| dog | 8 mo | p.o. | 0, 0.15, 0.4, 1.0 | 8 | 90 days | - |
| dog | 4.5-6 mo | p.o. | 0, 0.1, 0.4, 1.6 | 8 | 27 wk | - |
| | 4.5-6 mo | p.o. | 0, 0.4 | 4 | 27 wk | 3 wk |

* Ages at commencement of treatment.

** Each dosage group was composed of equal numbers of males and females.

Rat: 5-week study

There were no adverse effects related to treatment in the 0.1 and 0.4 g/kg groups.

By the end of the study, males in the 0.8 and 1.7 g/kg groups exhibited increased clotting times. An increase in serum alkaline phosphatase levels was seen in male rats in the 1.7 g/kg group. Histological changes in the stomach wall, similar to those seen in animals on 2.5 g/kg/day, were observed in males and females in the 1.7 g/kg group, and were considered to be primarily a consequence of the mechanical effects of drug accumulation.

In the high-dose (2.5 g/kg) group, all of the males were killed on day 9 and three of the females were killed or died later during the study. The deaths followed clinical deterioration due to accumulation of drug ester in the stomach. In affected rats there was thrombocytopenia in most cases and a slightly prolonged plasma-activated partial thromboplastin time in one case. Histological examination revealed mechanical damage to the stomach wall, and in one case there was scattered renal tubular eosinophilia and desquamation.

Rat: 90 day study

A number of rats died during the study; macroscopic and microscopic examination confirmed that these deaths were not related to cefuroxime axetil. The general condition of the surviving animals remained satisfactory throughout the study and the treatment did not affect the normal increase in body weight.

There was a slight decrease in total leukocyte counts observed in all dosage groups, resulting from a reduction in the number of neutrophils and lymphocytes and probably due to the protective effect of the test compound against those microbes that may influence leukocyte homeostasis.

A reversible decrease in plasma coagulation activity occurred in males, predominantly in the high-dose (1.0 g/kg) group. This may be explained by the direct action of cefuroxime axetil on the coagulation system or to reduced synthesis of coagulation factors as a result of suppression of Vitamin K-producing organisms in the intestine.

Rat: 28-week study

Rats in the 0.1 and 0.4 g/kg groups showed no significant toxicity and were in good general condition when the study was terminated after 62 to 65 days of treatment. However, the animals given 1.6 g/kg/day suffered from gastrointestinal trauma resulting from the mechanical effects of firm agglomerates of cefuroxime ester. Despite reduction of their dose to 1.0 g/kg/day on Day 7, they continued to deteriorate and either died or were killed after 10 to 14 days of treatment.

In a further 28 week study in rats, there were no deaths attributable to any toxic effect of cefuroxime axetil.

Apart from loose faeces, observed mainly during the first six weeks of the study in animals receiving 0.4 and 1.0 g/kg/day, there were no significant effects on the general condition of rats surviving to the end of the study. Salivation, extension of the forearms, and walking on the toes was seen in treated animals at the time of dosing, but were considered to be primarily a response to the dosing procedures rather than a toxic effect of the drug.

A reduction in leukocyte counts was observed in all treatment groups, probably reflecting a protective action of the antibiotic against minor infections. Other laboratory abnormalities observed included lengthened clotting times in males, reduction in SGOT and SGPT, and increases in serum transaminases in a few individual females without histological evidence of hepatic damage.

At the end of the treatment period, 48% of the animals in the high-dose (1.0 g/kg) had agglomerates of cefuroxime axetil in the stomach at autopsy, with one subject exhibiting an associated inflammation of the stomach wall.

Dog: 5-week study

Aside from a single case of vomiting shortly after dosing, the general condition of the dogs was not adversely affected by the treatment.

Laboratory abnormalities observed included transient decreases in total leukocyte and neutrophil counts, hyponatremia, and increased inorganic phosphorous and triglyceride levels. None of the individual results were sufficiently abnormal to be of clinical significance and there were no histological changes associated with treatment.

Dog: 90-day study

The general condition of the animals remained satisfactory throughout the study with normal increases in body weight attained. Isolated and occasional vomiting was the only apparent adverse effect, but this response can partly be attributed to the oral intubation.

Increases in erythrocyte sedimentation rate and in leukocyte and eosinophil counts were observed in animals with incidental helminthic infections. Females in the 0.4 and 1.0 g/kg groups exhibited increased total serum iron binding capacity. There was a statistically significant decrease in the absolute weights of livers in males and hearts in females in the 0.4 g/kg group, but this is insignificant when related to total body weights.

Dog: 27-week study

The general condition of the animals remained satisfactory throughout the study with the exception of 3 dogs, 2 of which were sacrificed suffering from illnesses unrelated to treatment.

In the high-dose (1.6 g/kg/day) group salivation and vomiting were noted, and in one dog there was a transient reduction in growth rate and a general loss of condition. Laboratory abnormalities included reduced erythrocyte counts, prolonged clotting times, reduction in plasma protein and cholesterol, and increase in plasma triglyceride. Post mortem examinations revealed no signs of organ toxicity.

Nephrotoxicity Studies

Single Dose Administration

Mouse

Mice received single subcutaneous doses of cefuroxime sodium (10 g/kg) alone or in combination with furosemide (50 mg/kg) or furosemide plus glycerol 5.4 mL/kg. Cefuroxime alone caused no nephrotoxicity; when administered concomitantly with furosemide there was proximal tubular necrosis in 2 out of 9 animals. The combination of furosemide and glycerol caused tubular necrosis in 5 of 8 animals but this was not influenced by the addition of cefuroxime.

Rat

Cefuroxime sodium at doses up to 10 g/kg was administered either alone or in combination with furosemide (100 mg/kg) or furosemide plus glycerol (3.15 mL/kg). Three of 6 animals exhibited proximal tubular necrosis in the inner cortex following administration of 4 g of cefuroxime alone. The incidence and severity of necrosis with increasing doses of cefuroxime. The incidence of tubular necrosis also increased when furosemide or furosemide plus glycerol were given concomitantly. Cefuroxime at 1g/kg enhanced the severity of the furosemide-glycerol-induced necrosis in the outer cortex. Treatment with furosemide plus glycerol also lowered the dosage of cefuroxime to 2 g/kg required to produce necrosis of the inner cortex.

Rat: Repeated dose study

Rats received cefuroxime at doses ranging from 1 to 5 g/kg/day subcutaneously for 10 days. There was no histological evidence of tubular necrosis at 5 g/kg, but transient increases in urine volume, protein and enzymes, which peaked at day 2-3 were observed. The body weights of the animals were significantly reduced for the high dose group.

Combination with aminoglycosides

Rats were treated with gentamicin (35 mg/kg) for 10 days. Cefuroxime sodium was given either concomitantly during the 10 days or as a single dose with the ninth dose of gentamicin. Gentamicin-induced tubular necrosis was not potentiated by the administration of single doses of cefuroxime up to 6 g/kg/day. Multiple doses of cefuroxime up to 4 g/kg protected rats against gentamicin-induced nephrotoxicity, but at doses of 6 g/kg/day of cefuroxime, severe tubular necrosis was observed after 4 days of treatment. Similar results were found with amikacin and tobramycin.

Mutagenicity Studies

Several standard assays were done to assess the mutagenic properties of cefuroxime axetil. These included both *in vitro* (Ames test, fluctuation test, gene conversion assay) and *in vivo* (micronucleus tests) assays.

***In vitro* assays**

Cefuroxime axetil was subjected to standard Ames tests, fluctuation tests, and gene conversion tests in concentrations of up to 208 µg/plate, 8.3 µg/mL, and 833 µg/mL respectively. The results of these tests were negative. Negative results were also obtained at high concentrations (833 µg/mL) in a modified fluctuation test in which the test strains were rendered resistant to cefuroxime's antibacterial properties. A weak, but statistically significant response was observed at 416 µg/mL, but this was not regarded as biologically significant since no effect was detected at 833 µg/mL.

***In vivo* micronucleus test**

Groups of five male mice received oral doses of cefuroxime axetil equivalent to 1.486, 1.114, 0.743 and 0.372 g/kg cefuroxime. Negative control groups received vehicle only and positive controls were dosed with 100 mg/kg cyclophosphamide. At either 24 or 48 h, groups of animals were killed and the bone marrow of both femurs collected. Smears were prepared and examined for micronuclei.

There was no significant increase in the proportion of polychromatic erythrocytes with micronuclei in any of the groups treated with cefuroxime axetil when compared with their negative control at either expression time.

The ratios of mature to immature erythrocytes observed in animals receiving cefuroxime axetil were not significantly different from the negative controls at either expression time.

Tolerance Studies

Cefuroxime axetil, applied as a 50% suspension in soft paraffin/liquid to intact and abraded guinea pig skin under occlusive dressing for 21 hours, produced negligible irritancy. Cefuroxime axetil was strongly sensitizing when applied to guinea pig skin. Sixteen days after application of sensitizing doses to 10 animals, challenge with the test material produced a positive erythematous response in 9 animals after 24 hours and in all 10 after 48 hours.

Reproduction and Teratology Studies:

Rodents

The reproductive toxicity of oral cefuroxime axetil was investigated in rats and mice as summarized in Table 9.

Table 9: Reproduction and Teratology Studies

| Animal | Sex | Doses (mg/kg/day) | Animal / Dose | Duration of Treatment | Significant Observations* |
|--------|-----|-------------------|---------------|--|--|
| mouse | F | 0, 150, 500, 1600 | 30** | day 7 to day 16 of pregnancy | - decreased number of implant(F ₀) - increased F ₁ male:female ratio |
| rat | F | 0, 125, 250, 500 | 20 | day 17 of pregnancy to day 21 <u>post partum</u> | -delayed pinna detachment (F ₁ females) |
| rat | M | 0, 125, 250, 500 | 10 | 70 days prior to mating | - delayed F ₁ mating, increased F ₂ |
| | F | 0, 125, 250, 500 | 30*** | 21 days before mating to day 21 <u>post partum</u> | - male: female ratio, delayed primary coat (F ₂ females), delayed eye opening (F ₂ males), delayed pinna detachment(F ₂) |
| rat | F | 0, 125, 250, 500 | 30*** | day 7 to day 16 of pregnancy | - decreased number of implants (F ₀), decreased number of live F ₁ fetuses |

*Apparent reproductive toxicity (i.e., other than F₀ organ toxicity) which was dose-related and not due to experimental artifacts or to the antimicrobial action of the drug (e.g. suppression of intestinal microflora).

**20 animals were killed at term; 10 were allowed to litter and complete the treatment.

***15 animals were killed at term; 15 were allowed to litter and complete the treatment.

The most common gross abnormality observed in offspring of treated dams was hydronephrosis, seen in comparable numbers at all dose levels including controls. There was no evidence that cefuroxime axetil had adversely affected fertility or peri-/post-natal development or organogenesis in rats or mice.

Rabbit

Rabbits were found to be unsuitable for reproductive toxicity testing of cefuroxime axetil. Six unmated females were treated with daily doses of 0.1 to 0.5 g/kg while 6 mated females received 0.2 g/kg. All but one animal showed a chronic loss of body weight and deterioration in overall conditions (3 animals died). One of the mated animals maintained a viable pregnancy. Two aborted and evidence of earlier resorption of implants was found in another two.

Post mortem examination of rabbits in both groups revealed liquefied intestinal contents and distended, gas-filled caecums in many cases. Changes in intestinal microflora were thought to be the cause of the observed toxicity.

REFERENCES OR SELECTED BIBLIOGRAPHY

1. Adams DH, Wood MJ, Farrell ID, Fox C, Ball AP: Oral cefuroxime axetil: clinical pharmacology and comparative dose studies in urinary tract infections. *J Antimicrob Chemother* 1985; 16:359-366.
2. Alvarez S, Jones M, Holtsclaw-Berk S, Guarderas J, Berk SL: In vitro susceptibilities and b-lactamase production of 53 clinical isolates of *Branhamella catarrhalis*. *Antimicrob Agents Chemother* 1985; 27:646-647.
3. Bluestone CD: Otitis media and sinusitis in children. Role of *Branhamella catarrhalis*. *rugs* 1986; 31(suppl 3):132-141.
4. Broekhuysen J, Deger F, Douchamps J, Freschi E, Mal N, Neve P, et al: Pharmacokinetic study of cefuroxime in the elderly. *Br J Clin Pharmacol* 1981; 12:801-805.
5. Brogden RN, Heel RC, Speight TM, Avery GS: Cefuroxime: a review of its antibacterial activity, pharmacological properties and therapeutic use.
6. Bundtzen RW, Toothaker RD, Nielson OS, Madsen PO, Welling PG, Craig WA: Pharmacokinetics of cefuroxime in normal and impaired renal function: comparison of high-pressure liquid chromatography and microbiology assays. *Antimicrob Ag Chemother* 1981; 19(3):443-449.
7. Chow AW, Bartlett KH: Comparative in vitro activity of ceftazidime (GR-20263) and other b-lactamase stable cephalosporins against anaerobic bacteria. *J Antimicrob Chemother* 1981; 8(suppl B):91-95.
8. Cooper TJ, Ladusans E, Williams PEO, Polychronopoulos V, Gaya H, Rudd RM: A comparison of oral cefuroxime axetil and oral amoxicillin in lower respiratory tract infections. *J Antimicrob Chemother* 1985; 16:373-378.
9. Curtis NAC, Orr D, Ross GW, Boulton MG: Affinities of penicillins and cephalosporins for the penicillin-binding proteins of *Escherichia coli* K-12 and their antibacterial activity. *Antimicrob Agents Chemother* 1979; 16:533-539.
10. Curtis NAC, Orr D, Ross GW, Boulton MG: Competition of b-lactam antibiotics for the penicillin-binding proteins of *Pseudomonas aeruginosa*, *Enterobacter cloacae*, *Klebsiella aerogenes*, *Proteus rettgeri*, and *Escherichia coli*. *Antimicrob Agents Chemother* 1979; 16:325-328.
11. Esposito S, Galante D, Barba D, Pennucci C, Limauro D: Correlation of b lactamase stability and antibacterial activity of b-lactams in b lactamaseproducing bacteria and respective transconjugants *Drugs Exptl Clin Res* 1986; 12:329-333.

12. Eykyn S, Jenkins C, King A, Phillips I: Antibacterial activity of cefuroxime, a new cephalosporin antibiotic, compared with that of cephaloridine, cephalothin, and cefamandole. *Antimicrob Ag Chemother* 1976; 9:690.
13. Fass RJ, Helsel VL: In vitro activity of U76,252(CS-807), a new oral cephalosporin. *Antimicrob Ag Chemother* 1988; 7:1082-1085.
14. Finn AL, Straughn A, Meyer M, Chubb J: Effect of dose and food on the bioavailability of cefuroxime axetil. *Biopharm Drug Dispos* 1987; 8:519-526.
15. Foord RD: Cefuroxime Human Pharmacokinetics. *Antimicrob Agents Chemother* 1976; 9:741-747.
16. Forsgren A, Walder M: Activity of common antibiotics against Branhamella catarrhalis, Haemophilus influenzae, pneumococci, Group A streptococci and Staphylococcus aureus in 1983. *Acta Otolaryngol (Stockh)* 1984; 407(suppl):43-49.
17. Garcia-Rodriguez JA, Garcia Sanchez JE, Garcia Garcia MI, Garcia Sanchez E, Munoz Bellido JL: In vitro activities of new oral b-lactams and macrolides against Campylobacter pylori. *Antimicrob Ag Chemother* 1989; 9:1650-1651.
18. Ginsburg CM, McCracken GH, Petryka M, Olser K: Pharmacokinetics and bactericidal activity of cefuroxime axetil. *Antimicrob Ag Chemother* 1985; 28(4):504-507.
19. Gold B, Rodriguez J: Cefuroxime: mechanisms of action, antimicrobial activity, pharmacokinetics, clinical applications, adverse reactions and therapeutic indications. *Pharmacother* 1983; 3(2):82-100.
20. Goldstein EJC and Citron DM: Comparative activities of cefuroxime, amoxicillin-clavulanic acid, ciprofloxacin, enoxacin, and ofloxacin against aerobic and anaerobic bacteria isolated from bite wounds. *Antimicrob Ag Chemother* 1988; 8:1143-1148.
21. Goto S: The *in vitro* and *in vivo* antibacterial activity of cefuroxime. *Proc R Soc Med* 1977; 70(suppl 9):56-62.
22. Grassi GG, Ferrara A, Grassi C: Comparative in vitro bactericidal activity of cephalosporins, in Spitzky KH, Karrer K (eds). *Proceedings of the Thirteenth International Congress of Chemotherapy*, Vienna, 1983:19-22.
23. Harding SM, Williams PEO, Ayrton J: Pharmacology of cefuroxime as 1-acetoxyethyl ester in volunteers. *Antimicrob Ag Chemother* 1984; 25(1):78-82.
24. Johnsson J, Brorson J-E: Influence of b-lactamase-producing strains of Branhamella catarrhalis and Haemophilus influenzae on certain b-lactam antibiotics. *J Antimicrob Chemother* 1983; 12:269-271.

25. Jones RN, Fuchs PC, Gavan TL, Gerlach EH, Barry AL, Thornsberry C: Cefuroxime, a new parenteral cephalosporin: collaborative in vitro susceptibility comparison with cephalothin against 5,887 clinical bacterial isolates. *Antimicrob Ag Chemother* 1977; 12(1):47-50.
26. Knapp CC, Washington JA, II: In vitro activities of LY163892, cefaclor, and cefuroxime. *Antimicrob Ag Chemother* 1988; 1:131-133.
27. Laverdiere M, Wheeler N, Sabath LD: Cefuroxime resistance to staphylococcal β -lactamases. *Proc R Soc Med* 1977; 70(suppl 9):72-73.
28. Liljequist BO, Gezelius L: In vitro activity of amoxicillin plus clavulanic acid against Haemophilus influenzae and Branhamella catarrhalis. *Eur J Clin Microbiol* 1986; 5:615-621.
29. Matthew M: Plasmid-mediated β -lactamases of gram-negative bacteria: Properties and distribution. *J Antimicrob Chemother* 1979; 5:349-358.
30. McCracken GH Jr, Ginsburg CM, Clahsen JC, Thomas ML: Pharmacologic evaluation of orally administered antibiotics and children: Effect of feeding on bioavailability. *J Pediatr* 1978; 62:738-743.
31. Neu HC, Fu KP: Cefuroxime, a β -lactamase-resistant cephalosporin with a broad spectrum of Gram-positive and -negative activity. *Antimicrob Ag Chemother* 1978; 13(4):657-664.
32. Neu HC, Saha G and Chin N-X: Comparative in vitro activity and β -lactamase stability of FK482, a new oral cephalosporin. *Antimicrob Ag Chemother* 1989; 10:1795-1800.
33. Neu HC: The emergence of bacterial resistance and its influence on empiric therapy. *Rev Infect Dis* 1983; 5(suppl):S9-S20.
34. Neu HC: Relation of structural properties of β -lactam antibiotics to antibacterial activity. *Am J Med* 1985; 79(2A):2-13.
35. O'Callaghan CH, Sykes RB, Griffiths A, Thornton JE: Cefuroxime, a new cephalosporin antibiotic: Activity *in-vitro*. (1976) *Antimicrobial Agents and Chemotherapy* 9:511-519.
36. Perez TE, Garcia Arenzana JM, Ayestaran I, Munoz Baroja I: Comparative activity in vitro of 16 antimicrobial agents against penicillin-susceptible meningococci and meningococci with diminished susceptibility to penicillin. *Antimicrob Ag Chemother* 1989; 9:1622-1623.
37. Philipson A, Stiernstedt G: Pharmacokinetics of cefuroxime in pregnancy. *Am J Obstet Gynecol* 1982; 142(7):823-828.

38. Slevin NJ, Aitken J, Thornley PE: Clinical and microbiological features of Branhamella catarrhalis bronchopulmonary infections. *Lancet* 1984;1:782-783.
39. Sommers DK, Van Wyk M, Moncrieff J: Influence of food and reduced gastric acidity on the bioavailability of bacampicillin and cefuroxime axetil. *Br J Clin Pharmacol* 1984; 18(4):535-539.
40. Sommers DK, Van Wyk M, Williams PEO, Harding SM: Pharmacokinetics and tolerance of cefuroxime axetil in volunteers during repeated dosing. *Antimicrob Ag Chemother* 1984; 25(3):344-347.
41. Stratton CW, Liu C and Weeks LS: Activity of LY146032 compared with that of methicillin, cefazolin, cefamandole, cefuroxime, ciprofloxacin, and vancomycin against Staphylococci as determined by kill-kinetic studies. *Antimicrob Ag Chemother* 1987; 8:1210-1215.
42. Sykes RB, Matthew M: The b-lactamases of gram-negative bacteria and their role in resistance to b-lactam antibiotics. *J Antimicrob Chemother* 1976; 2:115-157.
43. Tartaglione TA, Polk RE: Review of the new second-generation cephalosporins: cefonicid, ceforanide, and cefuroxime. *Drug Intell Clin Pharm* 1985; 19(3):188-198.
44. Thornsberry C, Biddle JW, Perine PL, Siegel M: In vitro susceptibility of b-Lactamase positive and b-Lactamase negative strains of Neisseria gonorrhoeae to cefuroxime. *Proc Roy Soc Med* 1977; Vol. 70; Supp. 9.
45. Van Landuyt HW, Pyckavet M: In vitro activity of cefotaxime against cephalothinresistant clinical isolates. *Antimicrob Agents Chemother* 1979; 16:109-111.
46. Williams PEO: Factors affecting the oral absorption of esterified antibiotics. *Biochem Soc Trans* 1985; 13:511-513.
47. Williams PEO, Harding SM: The absolute bioavailability of oral cefuroxime axetil in male and female volunteers after fasting and after food. *J Antimicrob Chemother* 1984; 13(2):191-196.
48. Wise R, Bennett SA, Dent J: The pharmacokinetics of orally absorbed cefuroxime compared with amoxicillin/clavulanic acid. *J Antimicrob Chemother* 1984; 13(6):603-610
49. A two-way, single-dose fasting bioavailability study of cefuroxime axetil 250 mg tablets in normal, healthy, non-smoking, male volunteers. September 1994.